## A Novel Porous Graphene Scaffold Prepared Using Freeze-Drying Technique for Orthopedic Approaches: Fabrication and Buckling Simulation Using GDQ Method

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Received: April 2020 Revised: June 2020 Accepted: July 2020

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**Abstract:** In this article, a novel bio-nanocomposite sample made of sodium alginate polymer, graphene nanosheets and wollastonite powder were produced using freeze-drying technique. The samples were mechanically and biologically evaluated using tensile strength and biological test. The phase and topological characterization were conducted by performing X-ray diffraction (XRD) and scanning electron microscopy (SEM) studies. Subsequently, using Euler-Bernoulli and Timoshenko beam (EBT and TBT) theories, the buckling response of the porous bionanocomposite soft tissue were analyzed with respect to graphene content. In order to solve the governing equations a sufficient numerical solution is proposed. Elastic modulus and mass density of the porous bionanocomposite are extracted from the experimental tests. The obtained results indicated the sample with 1 wt.% graphene sheet showed proper mechanical and biological features. Therefore, the sample with 1 wt.% graphene sheet can be used as potential case for light weight bone substitute applications.

Keywords: Scaffold, Graphene, Buckling analysis, Freeze-drying, Wollastonite.

## **1. INTRODUCTION**

Regarding human body disability of repairing bone defects, application of artificial bone tissue has received much attention in recent decades. New generation of bio-scaffolds are the appropriate approach for defective bone tissue. Bone defects are the consequences arising from trauma, accident, genetic or disease [1]. Recently, several studies have been conducted to produce novel porous bio-nanocomposite scaffold materials using various methods for clinical applications [2-4]. Biomaterials used in this category of application should be bioactive and made of synthetic or natural polymers, ceramics, and metals with sufficient porosity to ensure blood circulation inside human body using software and anatomical model [5-10]. Despite bioactivity, such materials have shown low mechanical properties and weak chemical beside problematic stability reaction in biological environment. Sufficient techniques have been presented to improve biomaterial mechanical characteristics such as compressive strength, fracture toughness, porosity and tensile strength [11-16]. Ceramic materials including hydroxyapatite (HA), biphasic calcium phosphate (CaP), bio-glasses (BG), and β-

 $(\beta TCP)$ tricalcium phosphate are being employed bone tissue engineering in applications [17-25]. Moreover, calcium silicates (CS), as a class of glass silicate, are one the bioactive-ceramic that has been utilized for orthopedic applications for decades [26-27]. The CS bioceramics are also used as coating cover for improving bioactivity of the porous scaffold implants [27]. Another fascinating property of CS is the ability to bond with natural bone inside the body, and the combination of CS with Magnesium (Mg) [28], Zinc (Zn), Strontium (Sr) [29] results in ameliorate molecular bonding properties for bone tissue approaches [30]. The CS bioceramic perform better mechanical behavior in comparison to other CaP bioceramics [31]. However, due to the required porosity and proper porous microarchitecture, the scaffold discloses unsuitable mechanical stiffness required for body weight and applied forces. Xu et al. [32] conducted in vivo bonegenerative resorption and evaluate the capacity of porous bioactive  $\beta$ -calcium silicate ( $\beta$ -CS) ceramics in a rabbit calvarial defect model. The β-CS ceramic was implanted in rabbit calvarial defect for 4, 8, and 16 weeks. Lin et al. [33] scrutinized the osteoporotic defect bone generation improvements by utilizing porous Sr-



substituted calcium ceramic scaffolds. Improving the mechanical characteristic of CS has been a controversial topic since the first usage of this material in biomedical application. Several strategies are proposed for improving its mechanical performance including reinforcement like graphene sheet (GS), magnetite nanoparticles (MNPs) or carbon (CNTs). Mehrali et al. nanotubes [34] synthesized a calcium silicate-reduced graphene oxide for hard tissue applications with enhanced mechanical properties using graphene reinforced GS and CS both indicated CS. proper biocompatibility features for bone microstructures like scaffolds and porous network. Shao et al. [35] developed a high strength Mg-doping wollastonite based scaffold by 3D printing technology having pore size and the heating procedure to strengthen the scaffold microstructure. Several polymers have been used for such purpose including chitosan, alginate, sodium alginate. gelatin application in pharmacokinetics [36-42, 53]. Xiong et al. [43] proposed a novel hybrid sodium alginate-based nanocomposite reinforce via graphene oxide and hydroxyapatite produced by freeze-drying method to mimic required mechanical and biological properties range. Another application of hybrid tissue is water treatment, for instance, Karkeh-abadi et al. [44] developed efficient adsorbents to remove radioactive pollutants from and industrial wastewater medical by functionalized CNT in sodium alginate-based nanocomposite. Lonita et al. [45] presented a novel study on the sodium alginate graphene oxide nanocomposite films with enhanced mechanical and thermal properties showing high interfacial adhesion between graphene and alginate matrix due to its hydrogen boding remarkably altered the mechanical properties and thermal stability of the bio-nanocomposite films. Scaffold fabrication techniques including solvent casting [46], particulate-leaching [47], gas foaming [48], phase separation [49], freezedrying [50-51], and 3D printing [52-53] are being used widely for biomedical usage. However, among these techniques, freeze-drying is one of the important, low cost, easily accessible and developed methods since it has the ability to control the porosity by the rate of freezing. Khorsand et al. [53] developed TiO<sub>2</sub> based scaffold doped in chitosan/HA utilizing

freeze drying technique with various hydroxyapatite ratios. The application bionanocomposites are extended in term of other fields studies including dental application [54-58]. In previous study, Foroutan et al. [59] concentrated on manufacturing polymer-based wollastonite-graphene scaffold with reinforcement. The mechanical and biological characterizations of prepared scaffold are studied. Moreover, the optimization is applied to the study for clear announcement of the optimum sample. Saber-Samandari et al. [60] prepared polyacrylamide-based nanocomposite scaffold incorporating nanohydroxyapatite by drying method. To evaluate the freeze mechanical behavior of the prepared bionanocomposite, the Timoshenko and Euler-Bernoulli beam theories are applied in different studies to investigate the buckling and bending behavior of beams. Sahmani et al. [61] investigated the nonlinear bending and instability of bredigite bioceramic incorporating magnetic nanoparticle. Foroutan et al. [62] proposed the buckling behavior analysis of current carrying nanowires regarding the magnetic field (MF) effects incorporating surface and nonlocal effects. Results revealed the MF increases the buckling load while nonlocal parameter reduces the buckling load. The biological feature mechanical and of biomaterials domain has been investigated with various techniques such as wet chemistry and mechanical activation methods. Several works have been conducted on drug loading and effect of nanoparticles on the composite. Molecular dynamic (MD) evaluation also can predict the mechanical properties of macron and micron scale [63-77]. Various numerical investigation has been performed on the composite to discover the influence of interphase of composite mathematically [68-74]. In the current paper, the sodium alginate biopolymer is composed with wollastonite ceramic to prepare proper bone orthopedic substitution for approaches. Moreover, GS is added in various content levels to the bio-nanocomposite. Mechanical and biological characterizations of the prepared bionanocomposite are surveyed. In order to investigate the mechanical behavior of the composed bio-nanocomposite, the Timoshenko and Euler-Bernoulli Beam theories (TBT) and (EBT) are used based upon the extracted elastic



modulus and mass density of the porous bionanocomposite from experimental examination to develop the classical beam responses.

## 2. EXPERIMENTAL PROCEDURES

### 2.1. Raw Materials

In the current study, the graphene nanosheet was used to enhance the new generation of calcium silicate (CS) scaffold. Sodium alginate (SA) as a natural polymer (Mw=216.12 g/mol, 98% purity, size <100 nm Merck company, Germany) were purchased. The wollastonite nanopowder was fabricated using high-energy ball milling (HEBM) and graphene sheet (GS) nanoplatelets (US Research Nanomaterials, Inc, +99.5%, 2.18 nm with 32 layers) are utilized.

### 2.2. Wollastonite Preparation

First, the preparation of wollastonite (WS) made of CaCO<sub>3</sub> and SiO<sub>2</sub> mixing by magnetic stirrer with 400 rpm and 25 °C dissolving in Acetic acid and water solution. Then the obtained gel is kept for 3 days in 70 °C. Furthermore, the temperature was increased to 150 °C for 24 h to dry the gel [59]. Dried gel is milled and filtered to reach particle size of 10-50 µm. Finally, two heating operation is applied on the particle in 700 °C and 1000 °C for 3 h to create the WS ceramic powder.

## 2.3. Nanocomposite Preparation

After the WS was prepared, manufacturing process of scaffold composites was performed. Briefly, 7.5 gr of sodium alginate powder was synthesized with 100 mL distilled water and 1 vol% acetic acid stirring for 3h and 1200 rpm with 50 °C. After that, 1.8 gr of WS powder were mixed with for various content of graphene (0, 1, 1)2, and 3 wt% GS) based on constant WS powder. In order to combine the prepared SA biopolymer and synthesized GS-WS powders the digital ultrasonic device is employed [59].

## 2.4. Scaffold Preparation

Then, the four porous bio-nanocomposite scaffolds were placed into freezer at -70 °C for 24 h. The frozen porous scaffold was set into the freeze-drier device for another 24 h at -45 °C with 0.01 bar to drain all the available waters to improvise required porosity (DORSA-TECH Company, Tehran, Iran) [59]. Porous scaffold removed from the freeze-drier machine after 48 h and placed in the incubator. Therewith, the

mechanical and biological tests are performed on the four porous bio-nanocomposites.

## 2.5. Characterization

## 2.5.1. Porosity Measurement

To evaluate the porosity of the bionanocomposite, the Archimedes and Image-J techniques were used. To this end, a graded cylindrical container with a fixed stilled water volume  $(V_1)$  is utilized [59]. Then, the prepared bio-nanocomposite was place into the water so that the water penetrates all the available porosity in the matter  $(V_2)$ . Afterward, the composite was pulled out of the container and the volume of water was measured  $(V_3)$ . The formula for the porosity percentage is written as equation 1:

Porosity (%) =  $(V_1 - V_3)/(V_2 - V_3) \times 100$ (1)

## 2.5.2. Tensile Strength Evaluation

The mechanical properties of the nanostructure including tensile strength and elastic modulus are investigated by the tensile strength device (SANTAM-STM50) by applying load of 0.5 mm/min [59]. The porous samples are cut into pieces with 9 mm diameter and 20 mm length corresponding to the fixed materials (WS-SA) and various content of GS powder. Obtained data are used to plot the stress-strain graph. The graph slop and the maximum value of stress are the elastic modulus and tensile strength, respectively.

## 2.6. Biological Evaluation

The Simulated Body Fluid (SBF) was utilized in order to evaluate the pH behavior of bionanocomposite. The ionic concentration of SBF is very similar to human blood plasma; therefore, it is used to evaluate the bio-nanocomposite pH behavior. To this end, the samples are drowned in SBF for 1, 3, 7, 14, 21 and 28-day's periods. The pH value is determined using a digital pH meter device at Amirkabir University Technology.

#### 2.7. Microstructural and Phase Characterization

The morphology and structural properties of Naalginate-WS-Graphene Sheet porous bionanocomposite scaffold are characterized through scanning electron microscopy (SEM) at Amirkabir University of technology (SERON Technology-AIS2100, South Korea, Display Magnitude 15 to 30000, Resolution 5/3 nm). The

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device voltage and electricity current were 40 KV and 30 mA, respectively. Moreover, phase composition of Na-alginate-WS-GS scaffold is investigated utilizing X-ray diffraction (XRD)

(INEL-EQUNIX 3000, French). Figure 1 displays the manufacturing process of fabricated scaffold bio-nanocomposite and required analysis of the structural characteristics.

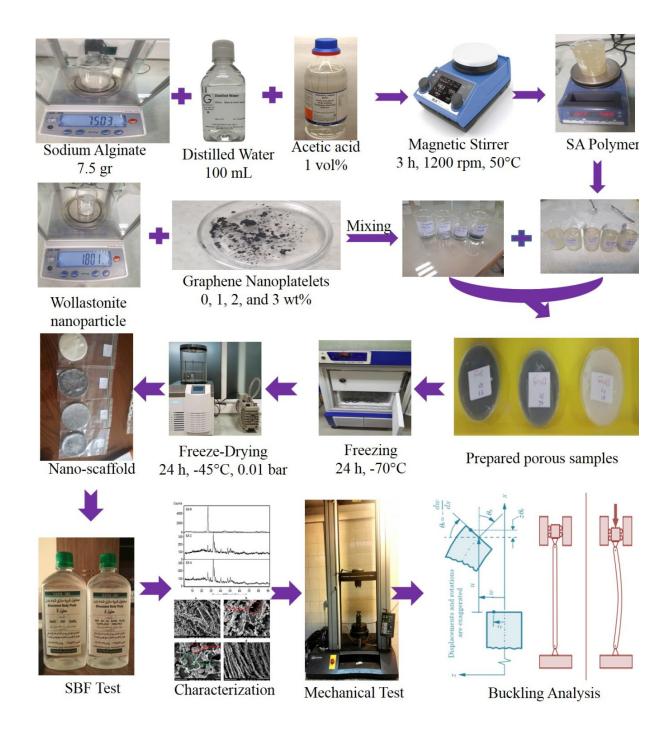


Fig. 1. Schematic of fabrication procedure of porous bio-nanocomposite made of Na-alginate-WS-reinforced with graphene oxide using freeze drying technique.



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## **2.8.** Analysis of Porous Scaffold Based on EBT and TBT Theories

In this section, the buckling behavior of bionanocomposite is investigated using TBT and EBT. Governing equations are derived based on both Euler-Bernoulli (EBT) and Timoshenko Beam Theories (TBT). Clamp-clamp (C-C) boundary conditions are imposed to simulate the bone replacement in the human body. Afterwards, the governing equations are solved Generalized Differential using Quadrature Method (GDQM) [12, 62 and 64]. According to Hamilton's principle the the buckling equilibrium equations is written as:

#### $\delta U - \delta W = 0$

Where  $\delta U$  and  $\delta W$  are virtual strain energy and virtual work done by external applied forces, respectively. Regarding the EBT and TBT theories, displacement fields are expressed as:

$$u(x,z) = u(x) - z \frac{dw(x)}{dx} + P(z) \left[ \frac{dw(x)}{dx} - \varphi(x) \right], \quad (2)$$
  
$$v(x,z) = 0,$$
  
$$w(x,z)w(x),$$

Where u, w, and  $\varphi$  are axial displacements, transverse displacements, and angle rotation of cross section about y-axis, respectively. P(z) for both EBT and TBT is defined as:

$$P(z)=0 EBT$$

$$P(z)=z TBT (3)$$

Based on defined displacement field, the strain components can be express as:

$$\varepsilon_{xx} = \frac{du}{dx} - P\frac{d\phi}{dx} - (z - P)\frac{d^2w}{dx^2},$$
(4)
$$\varepsilon_{xz} = \frac{1}{2}\frac{dP}{dz}\left(\frac{dw}{dx} - \phi\right)$$

In order to derive the classical governing equations based on Hamilton's principle, the stress resultants are defined as:

$$\left\{\hat{N}, \hat{M}, \hat{H}\right\} = \int_{A} \sigma_{xx} \left\{1, P, (z - P)\right\} dA, \hat{Q} = \int_{A} \sigma_{xz} \frac{dP}{dz} dA, (5)$$

Where  $\sigma_{xx}$  and  $\sigma_{xz}$  denote stress tensor components. Finally, the generalized governing equations (GGE) are derived according Hamilton's principle as follow:

$$\delta u : \frac{d\hat{N}}{dx} + f = 0, \qquad (6)$$
  
$$\delta w : \frac{d^{2}\hat{H}}{dx^{2}} + \frac{d\hat{Q}}{dx} + q - \frac{d}{dx} \left( N_{m} \frac{dw}{dx} \right) = 0, \qquad (6)$$
  
$$\delta \phi : \frac{d\hat{M}}{dx} - \hat{Q} = 0$$

Where  $\delta \phi$  is merely expressed in TBT. Moreover, q and f are the transverse and distributed axial loads, respectively. For EBT and TBT the derived equations based on Eq. 5 are written as:

$$\delta w : EI \frac{d^4 w}{dx^4} - N_m \frac{d^2 w}{dx^2} = 0,$$
 EBT (7)

$$\delta u : EA \frac{d^2 u}{dx^2} = 0, \tag{8}$$

$$\delta w : GA\left(\frac{d^2 w}{dx^2} - \frac{d^2}{dx^2}\right) - N_m \frac{d^2 w}{dx^2} = 0, \quad TBT$$
$$\delta \phi : -EI\frac{d^2 \phi}{dx^2} - GA\left(\frac{dw}{dx} - \right) = 0,$$

Where G is the shear modulus of elasticity that can be derived from G=E/(2(1+v)), v is the Poisson's ratio which can be derived from rule of mixture (ROM) as [65]:

$$v = v_{WS} V_{WS} + v_{SA} V_{SA} + v_{Gr} V_{Gr},$$

Where  $V_{ws}$ ,  $V_{SA}$ , and  $V_{Gr}$  are the volume fraction WS, SA, and GS, respectively. Moreover  $v_{Ws}$ ,  $v_{SA}$ , and  $v_{Gr}$  are the Poisson's ratios for WS, SA and GS, respectively. For acquiring more general responses the following dimensionless groups are utilized:

$$x^* = \frac{x}{L}, w^* = \frac{w}{L}, \phi^* = \phi, \beta = \frac{N_m L^2}{EI}$$
 EBT

$$\alpha = \frac{1}{AL^2}, \psi = \frac{G}{E}, \gamma = \frac{N_m}{EA} \qquad \text{TBT} \qquad (9)$$

# 2.8.1. Generalized Differential Quadrature *Method (GDQM)*

In order to solve the derived equation a numerical method is used. This method is known as Generalized Differential Quadrature Method (GDQM) [59-64] which is famous for its simple and low-cost calculations. Applying the boundary conditions is an outstanding point of this method since the boundary conditions can

DOI: 10.22068/ijmse.17.4.62

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be imposed simultaneously with the calculations. There are no restrictions on the number of mesh points applied for approximation in this method. Moreover, the weighted coefficients are derived by simple recurrence relation rather than a set of algebraic equations. These two advantages make GDQM a superior method in comparison to other DGM approaches [12, 62, 64, 65]. The partial derivations of function f with respect to spatial variables at the point  $x_i$  are expressed as:

$$\frac{d^{n}f}{dx^{n}} = \sum_{j=1}^{N} C_{ij}^{(n)} f(x_{j}), \qquad (10)$$

where  $C_{ij}^{(n)}$  and N are weighted coefficients matrix and the number of mesh points, respectively. Moreover, f can be replaced by u, w, and  $\varphi$ . For the first derivation, weighted coefficient matrix is written as:

$$C_{ij}^{(1)} = \left\{ \prod_{k=l,k\neq i,j}^{N} (x_{i-}x_{k}) \prod_{k=l,k\neq j}^{N} (x_{i-}x_{k}) \quad i \neq j \\ \sum_{k=l,k\neq i}^{N} \frac{1}{(x_{i-}x_{k})} \quad (i=j) \right\}$$
(11)

Where i, j are natural numbers. The weighted coefficient for higher-order derivation is expressed as:

$$C_{ij}^{(2)} = \sum_{k=1}^{N} C_{ik}^{(1)} C_{kj}^{(1)}$$

$$C_{ij}^{(3)} = \sum_{k=1}^{N} C_{ik}^{(2)} C_{kj}^{(1)}$$

$$C_{ij}^{(4)} = \sum_{k=1}^{N} C_{ik}^{(3)} C_{kj}^{(1)}$$
(12)

Another important factor in the accuracy and the convergence of the solution is the distribution pattern of the mesh points. Here, the Chebyshev-Gauss-Lobatto (CGL) distribution pattern is expressed as:

$$x_i = \frac{1}{2} \left\{ 1 - \cos \left[ \frac{(i-1)\pi}{(N-1)} \right] \right\}$$
 i=1, 2, 3,..., N

For improving the accuracy of the responses in implementation of GDQ method, the number of mesh points for a converged response are shown in Fig. 2.

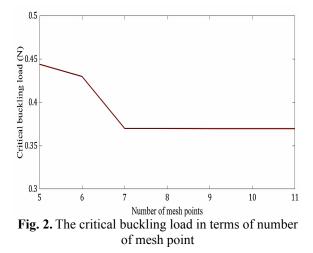
## 2.8.2. Applying the Boundary Conditions

The corrected collocation method [65] is applied to enforce the boundary conditions. For clampclamp boundary condition the following boundary conditions are imposed in the governing equations:

$$w(0)=w(1)=0$$
  
 $w'(0)=w'(1)=0$   
 $w(0)=w(1)=0$   
 $u(0)=u(1)=0$   
 $\phi(0)=\phi(1)=0$ 

Solving the equations based on GDQM and the essential boundary condition requires the application of eigenvalue theorem. After implementation of the eigenvalue theorem, the minimum dimensionless critical buckling load is obtained, then by utilizing the related dimensionless term the value of critical buckling load is acquired. The constant geometrical and mechanical values used in this research are assumed as:

L=15mm, D=1.5mm,  $v_{Ws}$ =0.3,  $v_{SA}$ =0.5,  $v_{Gr}$ =0.3



#### **3. RESULTS AND DISCUSSION**

In this section, the obtained results from the biological and mechanical tests are discussed and evaluated thoroughly. Moreover, XRD, SEM, and numerical analysis are debated regarding the obtained results. By the means of such tests and analysis the bio-nanocomposite behavior and bio-compatibility are examined, and the 1 wt% GS is selected as the most sufficient one.

#### **3.1. Mechanical Tests**

Fig. 3 depicts the stress-strain graph for four



levels of graphene weight percent. Extracted from the given graph, the elastic modulus is derived. As it is obvious from Fig. 3, increasing the GS weight percent from 0 wt% to 2 wt% would increase the material stiffness. It is absorbing that the GS percentage would decrease the material stiffness for the third level of graphene sheet (GS=3 wt%). This is probably happened due to the mixing features of the materials and agglomeration. Extracted data were obtained from previous work [59] to discover the elastic modulus of 0 to 3 wt% GS added to the sodium alginate-wollastonite.

## 3.2. pH Observation

In this section the pH behavior of prepared porous bony bio-nanocomposite is discussed. Fig. 4. represents the pH ranges for the basic (without GS) and optimum samples, and this pH rages can influence the rate of degradation. Since, WS is ceramic composed of silicate, it can affect the trend of pH changes and dominate the trend. Therefore, the basic sample can indicate the trend of pH changes for other various levels of GS powder. It means if the pH basic sample increases the other samples also follows the same trend. However, the GS can also influence the pH ranges. As it is shown in Fig. 4, the WS for pure sample increase the pH value, however; the graphene nanosheet is change reversely.

## 3.3. XRD Analysis

In this section, the XRD data are analyzed and discussed. Fig. 5 described the XRD reveals that produced bio-nanocomposite are containing GS since the XRD for the pure Gr pick on  $26.5^{\circ}$  20 are repeated for the bio-nanocomposite with 2 and 3 wt% GS. It is also discovered that the picks on approximate  $30^{\circ}$  are the one showed for the highest pick of WS on almost the same degree. The reasons of such small shifts on the degrees of highest peaks are inferred from the composite structure.

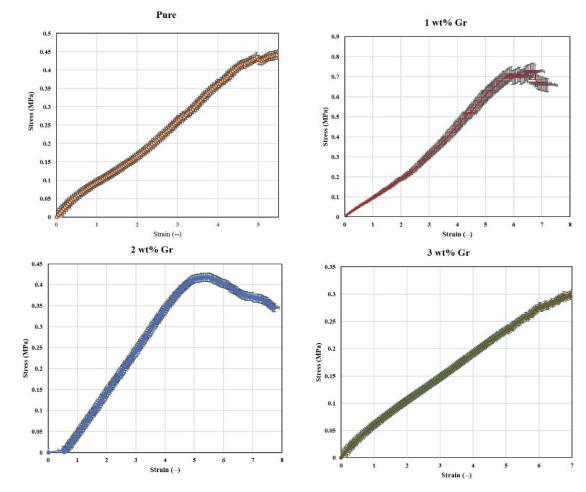


Fig. 3. Stain-stress graph for four levels of Graphene (i.e.; GS= 0-3 wt%)



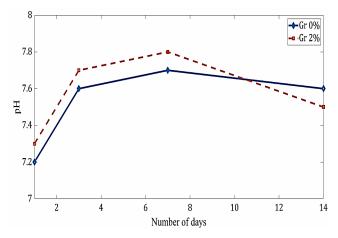


Fig. 4. The pH values in terms of number of days for basic and optimum composites

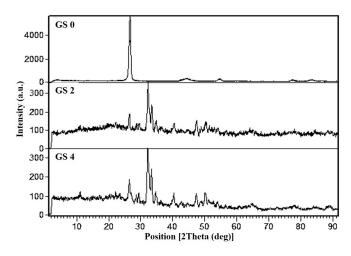


Fig. 5. XRD pattern of prepared scaffold using freeze drying technique containing various amount of GS

## 3.4. SEM Analysis

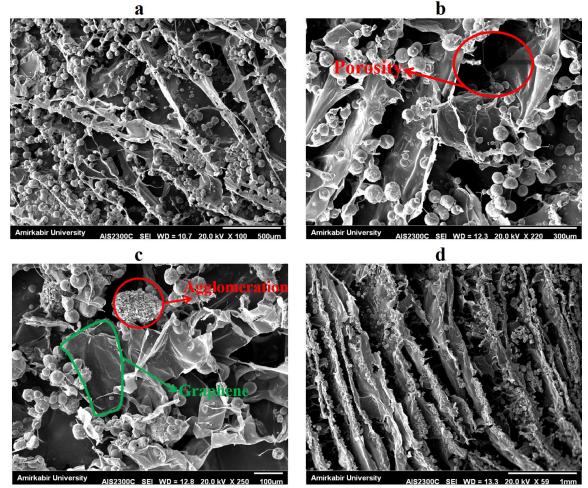
The SEM images is a way to see the composite structure including porosity, material, and samples layout. Fig. 6 shows the SEM images of four levels of GS weight percent. Fig. 6(a) shows when GS is not added to the bio-nanocomposite, the bonds between SA and WS are attended to be weak and string like. The WS nanoparticles are clustered and cause to create bio-nanocomposite with low porosity. Moreover, Fig. 6(b) shows the sample with 1 wt% GS chemical bonds between GS and WS are presented, thus the porosity size decrease to the rages of 150-200 µm. Fig. 6(c) displayed that when the GS weight percent increases, the agglomeration happens, and the GS agglomeration bond to the WS nanoparticles. This phenomenon decreases the porosity, but not happening all over the sample. Fig. 6(d) shows the porosities are shaped wall-like, and the connections between WS and GS homogenously distributed all over the sample.

#### 3.5. TBT and EBT Results

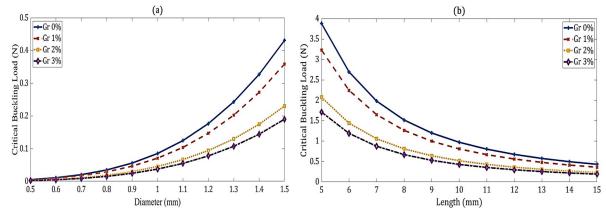
In this section, the buckling behavior of bionanocomposite beam restricted by clamp-clamp (C-C) boundary condition is discussed, and the obtained results are shown. Fig. 7 introduced the critical buckling load in terms of diameter and length are depicted for four levels of GS based on EBT. As it is previously shown, increasing the diameter incorporate escalations in the values of critical buckling load [12, 59, 62]. In contrast, the plays length of nanocomposite beam а contrariwise role which indicates reduction of critical buckling load as the length grow. Figure 8 plotted the critical buckling load as a function of diameter and length for various levels of GS. Fig. 7 and Fig. 8 displays the same results for length and diameter of bio-nanocomposite. Moreover, increasing the weight percentage of GS in the nanocomposite decreases the material stiffness and diminishes the critical buckling load [66 - 68].



Besides, a comparison of EBT and TBT value in the Figures discloses that the critical buckling load is lower in TBT because of considering more degrees of freedom. After evaluation of TBT and EBT, one can conclude that prepared scaffold compared to the other works have proper properties to load any drug and consider various nanoparticles in the sodium alginate and wollastonite matrix. Also, application of molecular dynamic (MD) can help the researchers to predict the mechanical properties of macron and micron scale without experimental testing [63-78].

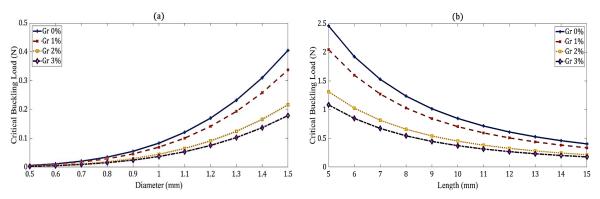


**Fig. 6.** SEM images of porous sample containing (a) 0 wt%, (b) 1 wt%, (c) 2 wt%, (d) 3 wt% of GS in the sodium alginate-wollastonite matrix fabricated using freeze drying technique



**Fig. 7.** Variation of critical buckling load in term of nanocomposite (a) diameter, and (b) length for four levels of Graphene based on EBT and clamp-clamp (C-C) boundary condition.





**Fig. 8.** Variation of critical buckling load in term of nanocomposite (a) diameter, and (b) length for four levels of Graphene based on TBT and clamp-clamp (C-C) boundary condition.

## 4. CONCLUSION

Elastic modulus and mass density of the bionanocomposite are extracted from the tests. experimental The obtained results indicated that the sample with 1 wt. % graphene nanosheet has shown proper mechanical and biological features. Therefore, the sample with 1 wt.% graphene can be used as potential case for light weight bone substitute applications. In this section, the obtained results from the biological and mechanical tests are discussed and evaluated thoroughly. Moreover, XRD, SEM, and numerical analysis are debated regarding the gained images and facts. Using such tests and analysis the bio-nanocomposite behavior and bio-compatibility were examined, and the 1 wt.% graphene nanosheet sample is selected as the most sufficient one. There are no restrictions on the number of mesh points applied for approximation in this method. Moreover, the weighted coefficients are derived by simple recurrence relation rather than a set of algebraic equations. These two advantages make GDQM a superior method in comparison to other DGM approaches. The synthesized calcium silicate-reduced graphene oxide for tissue applications hard with enhanced properties using mechanical graphene nanosheet reinforced CS was successfully achieved.

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